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# Native metals, electron bifurcation, and CO<sub>2</sub> reduction in early biochemical evolution

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Molecular hydrogen is an ancient source of energy and electrons. Anaerobic autotrophs that harness the H<sub>2</sub>/CO<sub>2</sub> redox couple harbour ancient biochemical traits that trace back to the universal common ancestor. Aspects of their physiology, including the abundance of transition metals, radical reaction mechanisms, and their main exergonic bioenergetic reactions, forge links between ancient microbes and geochemical reactions at hydrothermal vents. The midpoint potential of H<sub>2</sub> however requires anaerobes that reduce CO2 with H2 to use flavin based electron bifurcation - a mechanism to conserve energy as low potential reduced ferredoxins via soluble proteins - for CO<sub>2</sub> fixation. This presents a paradox. At the onset of biochemical evolution, before there were proteins, how was CO<sub>2</sub> reduced using H<sub>2</sub>? FeS minerals alone are probably not the solution, because biological CO2 reduction is a two electron reaction. Physiology can provide clues. Some acetogens and some methanogens can grow using native iron (Fe<sup>0</sup>) instead of H<sub>2</sub> as the electron donor. In the laboratory, Fe<sup>0</sup> efficiently reduces CO2 to acetate and methanol. Hydrothermal vents harbour awaruite, Ni<sub>3</sub>Fe, a natural compound of native metals. Native metals might have been the precursors of electron bifurcation in biochemical evolution.

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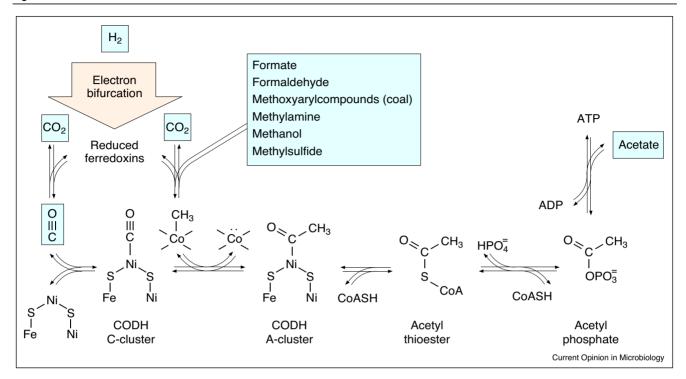
## Introduction

The only thing we know for sure about life's origin some 3.95 billion years ago [1] is that energy was required. Without energy release, no chemical reactions can take place that could ultimately lead to complex chemicals, metabolism or primitive ecosystems [2 $^{\circ}$ ]. Two sources of energy at origins are mainly discussed: ultraviolet (UV) light emitted from the sun [3,4] and chemical energy at hydrothermal vents [3,4]. Although UV light can be conveniently integrated into elegant laboratory syntheses of organic molecules at low temperatures and pressures [5,6], it does not connect to the microbial world (life) because no known form of microbial physiology is powered by UV light. From the biological standpoint, chemical energy at hydrothermal vents, in particular the  $H_2/CO_2$  redox couple, is interesting as a source of energy at origins. Why?

First, many forms of microbes use H<sub>2</sub> as a source of chemical energy for ATP synthesis in conjunction with a suitable electron acceptor such as CO<sub>2</sub> [7,8], and H<sub>2</sub>-dependent autotrophs provided the initial primary production that supported the first heterotrophic metabolisms [9]. Second, the Earth's crust has been generating large amounts of H<sub>2</sub> since there was liquid water, through a process called serpentinization [10]. In addition, biologists have long held that anaerobic autotrophs that reduce CO<sub>2</sub> using electrons from H<sub>2</sub> harbour the most ancient forms of metabolism [11-13]. H<sub>2</sub> dependent anaerobic autotrophs are furthermore rich in transition metal catalysts such as Fe and Ni [13], traits long regarded as ancient, and hydrogenases that extract the electron pair from H<sub>2</sub> to provide reduction equivalents and energy that drive metabolism forward [14].

Among the kinds of carbon and energy metabolism known among modern microbes, the acetyl-CoA pathway (or Wood-Ljungdahl pathway) as it is used in acetogens (bacteria) and methanogens (archaea) appears to be the most ancient [12]. It is the only exergonic pathway of biological CO<sub>2</sub> fixation known [12], all others require energy input in the form of ATP. The intermediate product of the pathway is a thioester, reactive compounds that have long been thought to play an important role in early chemical evolution [15]. In bacteria, the acetyl-CoA pathway generates acetyl phosphate from H<sub>2</sub>, CO<sub>2</sub> and phosphate (Figure 1). Acetyl phosphate is an excellent source of phosphorylation potential with a free energy of hydrolysis of -43 kJ/mol, 30% better than ATP. Even to

Figure 1



Scheme of energy conservation from ferredoxin to ATP in the acetyl CoA pathway. Redrawn after [12] incorporating intermediates in CO generation [13] and of substrate level phosphorylation [11]. Methanogenic growth on methoxy groups from coal was recently reported by [38\*]. The beige arrow indicates the requirement for electron bifurcation in the synthesis of low potential reduced ferredoxins with electrons from H<sub>2</sub> [7]. Insights into the mechanisms of electron bifurcation were recently revealed by the structures of two bifurcating enzymes [25\*,26\*]. Substrates and endproducts of the reversible reaction sequence are boxed in blue. The synthesis of the methyl group from H<sub>2</sub> and CO<sub>2</sub> entails energy investment, such that net synthesis of acyl phosphate or ATP from H<sub>2</sub> requires chemiosmotic coupling [6–8].

the level of the energy rich thioester the reaction is exergonic:

$$2CO_2 + 4H_2 + CoASH \rightarrow CH_3COSCoA + 3H_2O$$

$$\Delta G_{\rm o}' = -59 \,\mathrm{kJ/mol}[12].$$

Microbial genomes also point to the antiquity of anaerobic autotroph physiology. A recent study identified 355 ancient gene families that, based on their phylogenies, provided insights into the physiology and habitat of the last universal common ancestor, LUCA [16]. LUCA lived in a hot environment rich in gasses (H<sub>2</sub>, CO<sub>2</sub>, N<sub>2</sub>, CO) and metals, a habitat very similar to hydrothermal vents, which existed on the early Earth [10]. It used the acetyl-CoA pathway [16], its metabolism was rich in FeS and thioester dependent reactions, which are enriched in the ancient ATP-independent core of metabolism [17\*\*]. It harboured a diversified small molecule chemistry [18], many radical-based reactions that are dependent upon Sadenosyl methionine (SAM), which can form spontaneously in vitro [19°], and it harboured features found in microbes that today still inhabit ancient geochemical niches [20,21].

## The problem with H<sub>2</sub> and FeS

Despite its chemical simplicity, its abundance in ancient environments, and the clear tendency of H<sub>2</sub>-dependent autotrophs to branch deeply in phylogenetic trees [16], H<sub>2</sub> has a rather severe Achilles' heel as a source of electrons for CO<sub>2</sub> reduction at origins. Its midpoint potential is unfavourable for CO2 reduction beyond the near-equilibrium reaction with formate [22]. That is why microbes that reduce CO<sub>2</sub> with H<sub>2</sub> employ flavin based electron bifurcation [23], a biochemical mechanism that generates reduced ferredoxins (Fd<sup>-</sup>) with a midpoint potential on the order of -500 mV from H<sub>2</sub> with a more positive midpoint potential of only -414 mV [7]. Electrons from H<sub>2</sub> have to flow energetically uphill to low potential Fd. That might appear to violate the second law of thermodynamics, but electron bifurcation obeys the law in that one electron from H<sub>2</sub> is transferred energetically downhill to a high potential acceptor like NAD+ or heterodisulfide CoB-S-S-CoM [24], while the other is transferred uphill to Fd so that the overall energetics of the reaction are favourable [8].

Why is that problematic? The problem with  $H_2$  in an early evolution context is that flavin based electron bifurcation

is an elaborate physiological process that requires sophisticated proteins [25°,26°] working in concert with other proteins [7] as an energy metabolic pathway to reduce Fd (for reducing CO<sub>2</sub>). This presents a familiar chickenand-egg type paradox, namely how was CO<sub>2</sub> reduced with H<sub>2</sub> before there were proteins to catalyze electron bifurcation? One might counter that if early CO<sub>2</sub> fixation took place at hydrothermal vents, then electron bifurcation was not needed, because there was plenty of natural FeS around that could do the job of Fd<sup>-</sup> when it comes to fixing CO<sub>2</sub>.

But FeS minerals do not really solve the problem, because FeS has its own, different issues. FeS clusters in proteins are one electron donors, with iron undergoing Fe<sup>2+</sup> to Fe<sup>3+</sup> valence changes. The steps of biological CO<sub>2</sub> reduction in autotrophs are always two electron reactions [12]. In biology, the electrons from Fd<sup>-</sup> are donated to C in CO<sub>2</sub> via metals that readily undergo two electron reactions, such as Ni, Mo, or W atoms coordinated in proteins or cofactors (Figure 2), or electron pairs are donated via hydride transfer from organic cofactors like NAD(P)H [12].

Huber and Wächtershäuser [27] obtained excellent yields (40 mol%) of the thioester methyl thioacetate from CH<sub>3</sub>SH and CO using FeS, Ni<sup>2+</sup> and Fe<sup>2+</sup> salts, emulating the central anabolic reaction of the acetyl CoA pathway, but no one has reported genuine success involving FeS or other Fe<sup>2+</sup> species as a reductant for CO<sub>2</sub> in an early evolution context, electrochemical experiments where external voltage is applied aside [28]. Could it be that in the beginning, CO<sub>2</sub> was not reduced directly by H<sub>2</sub> at all? What does nature say?

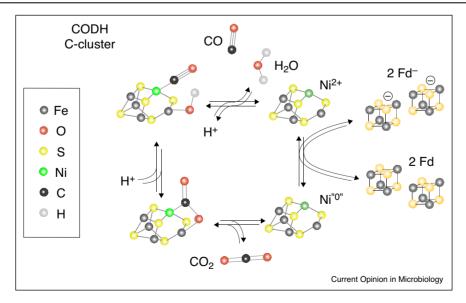
# Geochemical CO<sub>2</sub> reduction

Modern hydrothermal systems provide a check for CO<sub>2</sub> reduction in that CH<sub>4</sub> (ca. 1 mM) [29] and other small organic compounds including formate are present in hydrothermal effluent [30–32]. Those small organic compounds are not synthesized where vent effluent discharges into the ocean on the sea floor, rather, the organics are apparently made deep in the crust, as organics in terrestrial vents attest [33], CO<sub>2</sub> reduction in the modern crust is still not well understood in its details, although it is thought to stem from the same geochemical process that generates H<sub>2</sub> in hydrothermal effluent: serpentinization [30–32].

During serpentinization, H<sub>2</sub> synthesis stems from the reduction of water in hydrothermal systems via the oxidation of Fe<sup>2+</sup> that is present in the Earth's crust in vast amounts as iron magnesium silicates [10]. Fresh iron silicates for serpentinization are continuously supplied anew at spreading zones such as the mid-Atlantic ridge, where crust emerges as magma that subsequently cools, ultimately being recycled back into the mantle at subduction zones [34]. The isotope signatures of CH<sub>4</sub> emergent from hydrothermal systems are distinct from that of marine  $CO_2$  [29,32], the reasons for which are unclear. The rates of methane synthesis in laboratory scale serpentinization reactions so far are generally very slow [35°]. Thus, carbon is being reduced in serpentinizing systems, by a yet unidentified mechanism.

The process of serpentinization has been going on for the last 4.2 billion years, since there was liquid water on Earth [10]. Both its basic ingredients — water, Fe(II)-rich rocks,

Figure 2



Reaction mechanism of CO2 reduction in bifunctional CO dehydrogenase acetyl-CoA synthase (CODH/ACS) synthase in the acetyl-CoA pathway, proposed by Ragsdale [13], modified from [13]. C-cluster refers to the NiFeS cluster at the active site of the CODH enzyme [13].

#### Box 1 The early Earth in a nutshell

There is broad agreement among geochemists and planetary researchers that the early Earth was molten at some point, by the moon forming impact roughly 4.4 billion years ago, at the latest [10,33]. On the molten Earth (>1500 K), carbon that had been brought to the freshly accreted planet was converted to CO2, almost all of which was outgassed into the atmosphere, a small fraction being retained in magma oceans [10,33]. By about 4.4 Ga the magma oceans had cooled [33], by about 4.2 Ga there was liquid water on Earth [10], some outgassed from accretion and some delivered later by comets, and by around 4 Ga, perhaps as late as 3.7 Ga, the late heavy bombardment had come to an end [33]. By 3.95 Ga a carbon isotopic signature compatible with that produced by the acetyl-CoA pathway appeared [1].

As it relates to the source of energy for CO2 reduction, the relevant sequence of events is this: magma oceans in the molten phase oxidized Earth's early carbon to atmospheric CO2 and small amounts of mantle CO<sub>2</sub>. Because it cooled from magma, the primordial crust was depleted in water, consisted mostly of iron magnesium silicates with very low water content [10,55]. As the crust cooled, water condensed to surface oceans. Gravity pulled water into cracks in the solid crust, creating convective currents - the process of serpentinization set in. Serpentinization drew very CO2-rich water into the crust where serpentinization took place, such that H2 was synthesized in an otherwise H<sub>2</sub>-free environment and at sites where CO<sub>2</sub> existed in hydrothermal downcurrent water and as bound CO2 (carbonates) in a generally dry crust. The primordial interaction between H<sub>2</sub> and CO<sub>2</sub> thus probably took place deep in the crust, not at sites where hydrothermal vent effluent reached the ocean floor in contrast to earlier views [6]; H2 and CO2 first interacted in the presence of vast amounts of dry rock, and at temperatures likely exceeding 100 °C.

and heat for convection - and the chemical reaction are simple, as such the process appears to be occurring on the Saturn moon Enceladus as well [36°]. Serpentinization is a spontaneous process that releases chemical energy, a notable property that it shares with microbial energy metabolism. If serpentinization releases chemical energy, where does the energy that serpentinization releases come from? A look at early Earth history is instructive: the energy comes from the molten state of the early Earth (magma oxidizes carbon to CO<sub>2</sub>) and subsequent rockwater interactions, which then generate H2 in the presence of  $CO_2$  (Box 1).

Metagenomics tells us that approximately half of today's biomass lives in the crust in rocky, H<sub>2</sub> rich environments [37] and that substantial components of the modern subsurface biomass lives from the H<sub>2</sub>/CO<sub>2</sub> redox couple as acetogens and methanogens [38°], which fuel subsurface primary production. Today, microbes in the crust can also grow from organic carbon deposits such as coal [39\*\*], but that was not an option 4 billion years ago. Another main difference between today's crust and the primordial crust is that about half the water on Earth (i.e., roughly the volume of the ocean) is bound in the modern crust (and mantle), brought there by submarine hydrothermal activity [34]. The flipside of that coin is that the primordial oceans were twice as deep as today's [34], meaning that land for warm little ponds was probably in short supply.

# Whence electrons, if not H<sub>2</sub> or FeS?

Serpentinization alters the rocks that host hydrothermal activity. A very notable component of hydrothermally altered rocks is the mineral awaruite. Awaruite is an intermetallic compound with the formula Ni<sub>3</sub>Fe (or Ni<sub>2-3</sub>Fe): native transition metals with oxidation state zero. It is a normal constituent of serpentinizing hydrothermal systems [35°,40], formed there naturally during serpentinization under conditions where high H<sub>2</sub> activities of  $\sim$ 200 mmol/kg [41] reduce the divalent metal ions.

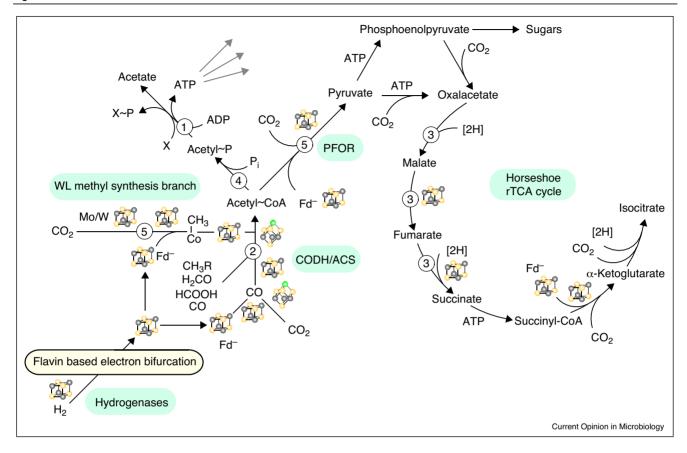
Almost 20 years ago Horita and Berndt [42] reported that Ni<sub>3</sub>Fe could catalyze the synthesis of methane in mmol/ kg amounts from H<sub>2</sub> and CO<sub>2</sub> [42] in simulated hydrothermal conditions (200–400 °C, ca. 50 MPa) although it cannot be excluded that awaruite was the reductant rather than the catalyst, at least in part. They also reported that after 1-2 weeks at lower temperature (200 °C) carbon compounds of oxidation state intermediate between CO<sub>2</sub> and CH4 were obtained in amounts similar to or exceeding those observed for CH<sub>4</sub>. Guan et al. [43] showed that Fe<sup>0</sup> in the presence of salts will reduce CO<sub>2</sub> to CH<sub>4</sub>, C<sub>3</sub>H<sub>8</sub>, CH<sub>3</sub>OH and C<sub>2</sub>H<sub>5</sub>OH in the 10-70 μM range at room temperature. He et al. [44] reported reduction of CO<sub>2</sub> to formate and acetate in the 1–10 mM range using nanoparticular Fe<sup>0</sup> at 80–200 °C. Moreover, Muchowska et al. [45°] recently showed native iron to accelerate and promote reactions of the reverse citric acid cycle (Figure 3). The message is that native metals can efficiently reduce CO<sub>2</sub>. That is not the case for either H<sub>2</sub> alone or for FeS minerals. The native metals are interesting.

Awaruite is today synthesized where H<sub>2</sub> is produced and where organic compounds are being made. Are native metals involved in geochemical CO<sub>2</sub> reduction, and were they involved in primordial CO<sub>2</sub> reduction? Considering the mechanism proposed by Steve Ragsdale [13] for acetyl-CoA synthesis in acetogens (Figure 2), the active Ni species for CO<sub>2</sub> reduction in the (4 billion year old) biochemical reaction is formally Ni<sup>0</sup>. The electrons are delivered to the enzyme one at a time via FeS clusters in Fd<sup>-</sup>, but are delivered to carbon as a pair. The Fe<sup>2+</sup> species in FeS clusters perform one electron chemistry, but the two electron carbon reduction reaction is performed by the transiently native metal. This might be a clue about ancient life.

## Microbes always have the last word

What do anaerobic microbes say about native metals? Basically they say 'yes, please', and they are the source of much corrosion to things made of steel, as recently surveyed by Enning and Garrelfs [46]. Early reports showed that methanogens grow from Fe<sup>0</sup> and CO<sub>2</sub> [47], but questions remained whether the growth was really from iron or just from H<sub>2</sub> generated by interaction of iron with water. Dinh et al. [48] then showed that methanogens grew rapidly on native iron as an electron

Figure 3



The acetyl-CoA pathway and the incomplete reverse citric acid cycle as the core carbon end energy metabolism in some modern microbes and the first microbes, modified from Ref. [6] to include the energetic impediment imposed by flavin based electron bifurcation in ferredoxin reduction with electrons from H<sub>2</sub> [7] and updated to include new information. Participation of FeS or FeNiS clusters is indicated using database information for the corresponding enzymes indicated in Fuchs [12]. Note the paucity of ATP-dependent steps and the acyl phosphate/ATP-generating steps, indicated with ATP next to the reaction (see also Ref. [17\*\*]). Circled numbers at reactions indicate that the non-enzymatic laboratory reaction has been reported as follows. (1) Kitani et al. reported that ATP is readily generated from acetyl phosphate and ADP using only Fe3+ [57] or Fe2+ [58] as catalysts. (2) Wächtershäuser and Huber reported the divalent metal ion-catalyzed synthesis of acetyl thioesters from CH<sub>2</sub>SH and CO [26\*]. (3) Muchowska et al. showed these steps to proceed spontaneously in vitro using native metal and metal ion catalysts [45\*\*]. (4) Weber [56] reported the synthesis of pyrophosphate from an acetylthioester and P<sub>i</sub>, whereby the presence of acetyl phosphate was inferred but not directly shown [6]. (5) Varma et al. [54\*\*] recently demonstrated synthesis of metal bound methyl groups and synthesis of pyruvate from CO<sub>2</sub> with native metals as the source of electrons. The free energy of hydrolysis for acetyl phosphate (-43 kJ/mol) is greater than that for ATP (-31 kJ/mol) [6]. In the context of early biochemical evolution, reactions that are today coupled to ATP hydrolysis would be thermodynamically even more favourable if coupled to acetyl phosphate (or other acyl phosphate) hydrolysis, an argument in favour of acetyl phosphate as a primitive energy currency [6,59]. The involvement of reductants other than Fd in the enzymatic reaction is indicated with [2H]. Abbreviations: CODH/ACS, carbon monoxide dehydrogenase/acetyl-CoA synthase; WL, Wood-Ljungdahl; PFOR, pyruvate ferredoxin oxidoreductase.

source, suggesting that H<sub>2</sub> was not involved as an intermediate. More recently, Tan et al. [49] showed that Methanosarcina barkeri will grow on Fe<sup>0</sup> as an electron source, but only when deprived of standard electron donors such as CH<sub>3</sub>OH or H<sub>2</sub>.

The molecular mechanisms of microbial electron extraction from Fe<sup>0</sup> are so far elusive. Lohner *et al.* [50] reported Fe<sup>0</sup> oxidation by methanogens in the presence of externally applied voltage. The results suggested that methanogens can access electrons from Fe<sup>0</sup> via routes that do not involve H<sub>2</sub>. More recent results by Deutzmann and colleagues using applied potentials suggest that methanogens might acquire electrons from Fe<sup>0</sup> via extracellular enzymes that oxidize the native metal to standard electron sources such as  $H_2$  or formate [51,52]. There are many reports about anaerobic growth on iron, many involving methanogens or sulfate reducers [46]. But acetogens?

Kato et al. [53°] recently isolated acetogens from the genus Sporomusa (firmicutes) that grow on native iron without externally applied potentials. Most acetogens that Kato et al. [53°] tested do not grow on iron as the electron source, indicating the presence of genetically specified mechanisms to access electrons from Fe<sup>0</sup>, in line with the conclusions of Dinh et al. [48] for sulfate reducers and methanogens. For sulfate reducers, the terminal acceptor of electrons from Fe<sup>0</sup> can be a sulfur compound or CO<sub>2</sub>, for acetogens and methanogens, the terminal acceptor is CO<sub>2</sub>. The proteins, cofactors and mechanisms involved in Fe<sup>o</sup> oxidation are still unknown [46.52.53°]. It is possible that non-enzymatic reactions of Fe<sup>0</sup> with CO<sub>2</sub> such as those generating formate, methanol, and acetate in the laboratory [43,44] play a role in microbial growth on iron. Very recent work reports the non-enzymatic synthesis of pyruvate from aqueous CO<sub>2</sub> and Fe<sup>0</sup> under mild hydrothermal conditions [54°°].

## **Conclusions**

As usual, nature leaves us with observations and questions. Is the oxidation of native iron (and other metals) an ancient trait, preserved from the very earliest phases of biological CO<sub>2</sub> reduction and is it prevalent in hydrothermal vents, where Ni<sub>3</sub>Fe is still made today? The two electron iron oxidation reaction  $Fe^0 \rightarrow Fe^{2+} + 2e^-$  has a midpoint potential of  $E_o' = -470 \text{ mV } [53^{\circ}]$ , more negative than that for hydrogen  $H_2 \rightarrow 2H^+ + 2e^-$ , with  $E_0' = -$ 410 mV. Might anaerobic autotrophs that oxidize Fe<sup>0</sup> short circuit flavin based electron bifurcation to generate their low potential reduced ferredoxins? If so, they still would have to direct two electron (Fe<sup>0</sup>) to one electron (FeS clusters in Fd<sup>-</sup>) reactions. The physiological reactions by which microbes access Fe<sup>0</sup> and other native metals as electron sources might uncover hints about early life, possibly even probing a phase of physiological evolution before there was genetically encoded electron bifurcation.

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## References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- •• of outstanding interest
- Tashiro T, Ishida A, Hori M, Igisu M, Koike M, Méjean P, Takahata N, Sano Y, Komiya T: Early trace of life from 3.95 Ga sedimentary rocks in Labrador, Canada. Nature 2017, 549:516-
- Judson OP: The energy expansions of evolution. Nat Ecol Evol 2017, 1 0138.

A concise and interesting overview of how life accessed new sources of energy during evolution.

- Patel BH, Percivalle C, Ritson DJ, Duffy CD, Sutherland JD: Common origins of RNA, protein and lipid precursors in a cyanosulfidic protometabolism. Nat Chem 2015, 7:301-307.
- Bonfio C, Valer L, Scintilla S, Shah S, Evans DJ, Jin L, Szostak JW, Sasselov DD, Sutherland JD, Mansy SS: UV-light-driven prebiotic synthesis of iron-sulfur clusters. Nat Chem 2017, **60**:2817.

- Baross JA, Hoffman SE: Submarine hydrothermal vents and associated gradient environments as sites for the origin and evolution of life. Orig Life Evol Biosph 1985, 15:327-345
- Martin W, Russell MJ: On the origin of biochemistry at an alkaline hydrothermal vent. Philos Trans R Soc Lond B: Biol Sci 2007 362-1887-1925
- Basen M, Müller V: "Hot" acetogenesis. Extremophiles 2017, **21**:15-26.
- Buckel W, Thauer RK: Energy conservation via electron bifurcating ferredoxin reduction and proton/Na(+) translocating ferredoxin oxidation. Biochim Biophys Acta 2013,
- Schönheit P, Buckel W, Martin W: On the origin of heterotrophy. Trends Microbiol 2016, 24:12-25.
- Sleep NH, Bird DK, Pope EC: Serpentinite and the dawn of life. Philos Trans R Soc Lond B: Biol Sci 2011, 366:2857-2869
- 11. Decker K, Jungermann K, Thauer RK: Energy production in anaerobic organisms. Angew Chem Int Ed Engl 1970, 9:138-158.
- Fuchs G: Alternative pathways of carbon dioxide fixation: insights into the early evolution of life? Annu Rev Microbiol 2011, 65:631-658.
- Ragsdale SW: Nickel-based enzyme systems. J Biol Chem 2009, 284:18571-18575.
- Søndergaard D, Pedersen CNS, Greening C: HydDB: a web tool for hydrogenase classification and analysis. Sci Rep 2016,
- Semenov SN, Kraft LJ, Ainla A, Zhao M, Baghbanzadeh M, Campbell VE, Kang K, Fox JM, Whitesides GM: Autocatalytic, bistable, oscillatory networks of biologically relevant organic reactions. Nature 2016, 537:656-660.
- 16. Weiss MC, Sousa FL, Mrnjavac N, Neukirchen S, Roettger M, Nelson-Sathi S, Martin WF: The physiology and habitat of the last universal common ancestor. Nat Microbiol 2016, 1:16116.
- 17. Goldford JE, Hartman H, Smith TF, Segrè D: Remnants of an ancient metabolism without phosphate. Cell 2017, 168:1126-

A comprehensive investigation of biochemical pathways that uncovers an ancient core of ATP-independent and phosphate-independent reactions in modern metabolism.

- 18. Michael AJ: Evolution of biosynthetic diversity. Biochem J 2017,
- 19. Laurino P, Tawfik DS: Spontaneous emergence of Sadenosylmethionine and the evolution of methylation. Angew Chem Int Ed Engl 2017, 56:343-345.

Shows that S-adenosylmethionine (SAM), an ancient and conserved cofactor that is central to the biosynthesis of other cofactors, can form from simpler components without enzymes.

- Giovannelli D, Sievert SM, Hügler M, Markert S, Becher D, Schweder T, Vetriani C: Insight into the evolution of microbial metabolism from the deep-branching bacterium, Thermovibrio ammonificans. eLife 2017, 6:2050.
- 21. Nayak DD, Mahanta N, Mitchell DA, Metcalf WW: Posttranslational thioamidation of methyl-coenzyme M reductase, a key enzyme in methanogenic and methanotrophic Archaea. eLife 2017, 6:108.
- 22. Schuchmann K, Müller V: Direct and reversible hydrogenation of CO<sub>2</sub> to formate by a bacterial carbon dioxide reductase. Science 2013. 342:1382-1385.
- 23. Herrmann G, Jayamani E, Mai G, Buckel W: Energy conservation via electron-transferring flavoprotein in anaerobic bacteria. J Bacteriol 2008. 190:784-791.
- 24. Kaster A-K, Moll J, Parey K, Thauer RK: Coupling of ferredoxin and heterodisulfide reduction via electron bifurcation in hydrogenotrophic methanogenic archaea. Proc Natl Acad Sci USA 2011, 108:2981-2986.
- 25. Lubner CE, Jennings DP, Mulder DW, Schut GJ, Zadvornyy OA, Hoben JP, Tokmina-Lukaszewska M, Berry L, Nguyen DM,

Lipscomb GL et al.: Mechanistic insights into energy conservation by flavin-based electron bifurcation. Nat Chem Biol 2017, 13:655-659.

Provides important insights into the mechanistic basis of flavin based electron bifurcation.

- 26. Wagner T, Koch J, Ermler U, Shima S: Methanogenic
- heterodisulfide reductase (HdrABC-MvhAGD) uses two noncubane [4Fe-4S] clusters for reduction. Science 2017, **357**:699-703

Provides important insights into the mechanistic basis of flavin based electron bifurcation.

- 27. Huber C. Wächtershäuser G: Activated acetic acid by carbon fixation on (Fe, Ni)S under primordial conditions. Science 1997,
- 28. Roldan A, Hollingsworth N, Roffey A, Islam H-U, Goodall JBM, Catlow CRA, Darr JA, Bras W, Sankar G, Holt KB et al.: Bioinspired CO<sub>2</sub> conversion by iron sulfide catalysts under sustainable conditions. Chem Commun (Camb) 2015, 51:7501-
- Proskurowski G, Lilley MD, Seewald JS, Früh-Green GL, Olson EJ, Lupton JE, Sylva SP, Kelley DS: **Abiogenic hydrocarbon** production at lost city hydrothermal field. Science 2008, 319:604-607.
- 30. Schrenk MO, Brazelton WJ, Lang SQ: Serpentinization, carbon, and deep life. Rev Miner Geochem 2013, 75:575-606.
- McCollom TM, Seewald JS: Serpentinites, hydrogen, and life. Elements 2013, 9:129-134.
- 32. McDermott JM, Seewald JS, German CR, Sylva SP: Pathways for abiotic organic synthesis at submarine hydrothermal fields. *Proc Natl Acad Sci USA* 2015, **112**:7668-7672.
- Etiope G, Schoell M: Abiotic gas: atypical, but not rare. Elements 2014, 10:291-296.
- Zahnle K, Arndt N, Cockell C, Halliday A, Nisbet E, Selsis F, Sleep NH: Emergence of a habitable planet. Geology and Habitability of Terrestrial Planets. Springer; 2007:35-78.
- McCollom TM: Abiotic methane formation during experimental serpentinization of olivine. Proc Natl Acad Sci USA 2016, **113**:13965-13970

An important investigation into the processes of serpentinization and its evolutionary significance.

Waite JH, Glein CR, Perryman RS, Teolis BD, Magee BA, Miller G, Grimes J, Perry ME, Miller KE, Bouquet A et al.: Cassini finds molecular hydrogen in the Enceladus plume: evidence for hydrothermal processes. Science 2017, 356:155-159.

Reports evidence for the existence of serpentinization on the Saturn moon Enceladus, which has a rocky core and liquid water.

- 37. Whitman WB, Coleman DC, Wiebe WJ: Prokaryotes: the unseen majority. Proc Natl Acad Sci USA 1998, 95:6578-6583.
- Colman DR, Poudel S, Stamps BW, Boyd ES, Spear JR: The deep, hot biosphere: twenty-five years of retrospection. *Proc Natl Acad Sci USA* 2017, **114**:6895-6903.

A timely perspective on microbial communities in the deep biosphere and Thomas Gold's ideas on the origin of fossil fuels on Earth.

Mayumi D, Mochimaru H, Tamaki H, Yamamoto K, Yoshioka H, Suzuki Y, Kamagata Y, Sakata S: Methane production from coal by a single methanogen. Science 2016, 354:222-225

Reports growth of a methanogen on the arylmethoxygroups in coal.

- 40. Schwarzenbach EM, Gazel E, Caddick MJ: Hydrothermal processes in partially serpentinized peridotites from Costa Rica: evidence from native copper and complex sulfide assemblages. Contrib Miner Petrol 2014, 168:1079.
- 41. Klein F, Bach W: Fe-Ni-Co-O-S phase relations in peridotiteseawater interactions. J Petrol 2009, 50:37-59.

- 42. Horita J, Berndt ME: Abiogenic methane formation and isotopic fractionation under hydrothermal conditions. Science 1999,
- 43. Guan G. Kida T. Ma T. Kimura K. Abe E. Yoshida A: Reduction of aqueous CO<sub>2</sub> at ambient temperature using zero-valent ironbased composites. Green Chem 2003, 5:630-634.
- 44. He C, Tian G, Liu Z, Feng S: A mild hydrothermal route to fix carbon dioxide to simple carboxylic acids. Org Lett 2010, 12:649-651.
- 45. Muchowska KB, Varma SJ, Chevallot-Beroux E, Lethuillier-Karl L, Li G, Moran J: Metals promote sequences of the reverse Krebs cycle. Nat Ecol Evol 2017, 1:1716-1721.

An important investigation showing that reactions of the citric acid cycle take place without enzymes, catalyzed by metals and metal ions.

- 46. Enning D, Garrelfs J: Corrosion of iron by sulfate-reducing bacteria: new views of an old problem. Appl Env Microbiol 2014, 80:1226-1236.
- 47. Daniels L, Belay N, Rajagopal BS, Weimer PJ: Bacterial methanogenesis and growth from CO<sub>2</sub> with elemental iron as the sole source of electrons. Science 1987, 237:509-511.
- 48. Dinh HT, Kuever J, Mußmann M, Hassel AW, Stratmann M, Widdel F: Iron corrosion by novel anaerobic microorganisms. Nature 2004, 427:829-832.
- 49. Tan JL, Goh PC, Blackwood DJ: Influence of H2S-producing chemical species in culture medium and energy source starvation on carbon steel corrosion caused by methanogens. Corros Sci 2017, 119:102-111.
- Lohner ST, Deutzmann JS, Logan BE, Leigh J, Spormann AM: Hydrogenase-independent uptake and metabolism of electrons by the archaeon Methanococcus maripaludis. ISME J 2014. 8:1673-1681.
- 51. Deutzmann JS, Spormann AM: Enhanced microbial electrosynthesis by using defined co-cultures. ISME J 2016, **11**:704-714
- 52. Deutzmann JS, Sahinb M, Spormann AM: Extracellular enzymes facilitate electron uptake in biocorrosion and bioelectrosynthesis. mBio 2015, 6:e00496-e515.
- 53. Kato S, Yumoto I, Kamagata Y: Isolation of acetogenic bacteria that induce biocorrosion by utilizing metallic iron as the sole electron donor. Appl Env Microbiol 2015, 81:67-73.

Reports growth of acetogens using native iron as the source of electrons for CO<sub>2</sub> reduction.

54. Varma SJ, Muchowska KB, Chatelain P, Moran J: Metals enable a non-enzymatic acetyl CoA pathway. bioRxiv 2017 http://dx.doi. org/10.1101/235523

Reports the synthesis of formate, methanol, acetate, and pyruvate (10-100 micromolar range) from CO<sub>2</sub> and native metals under mild aqueous conditions.

- Arndt N, Nisbet E: Processes on the young Earth and the habitats of early life. Annu Rev Earth Planet Sci 2012, 40:521-549.
- 56. Weber AL: Formation of pyrophosphate, tripolyphosphate, and phosphorylimidazole with the thioester, N,S diacetyl-cysteamine, as the condensing agent. J Mol Evol 1981, 18:24-29.
- 57. Kitani A, Tsunetsugu S, Suzuki A, Ito S, Sasaki K: Fe(III)-ioncatalysed non-enzymatic transformation of ADP into ATP. Chem Soc Perkin Trans 1991, 3:323-329.
- 58. Kitani A, Tsunetsugu S, Suzuki A, Ito S, Sasaki K: Fe(III)-ioncatalysed non-enzymatic transformation of adenosine diphosphate into adenosine triphosphate part II. Evidence of catalytic nature of Fe ions. Bioelectrochem Bioenerget 1995,
- 59. Martin WF, Thauer RK: Energy in ancient metabolism. Cell 2017, **169**:953-955.